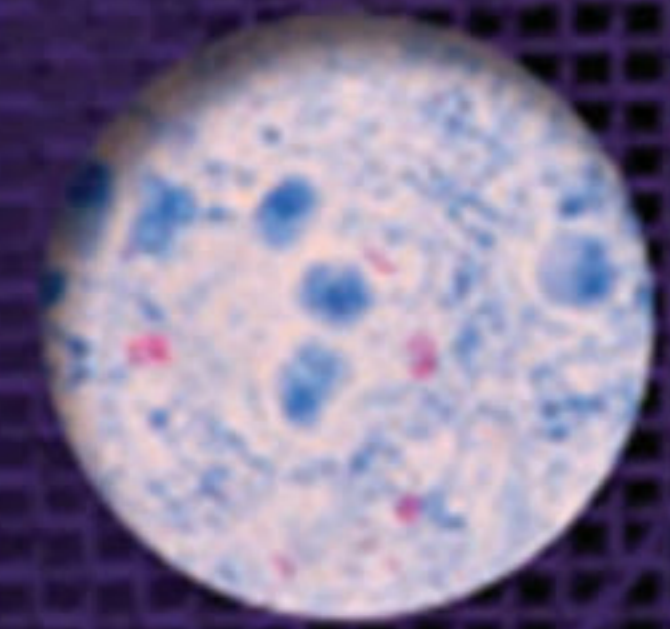
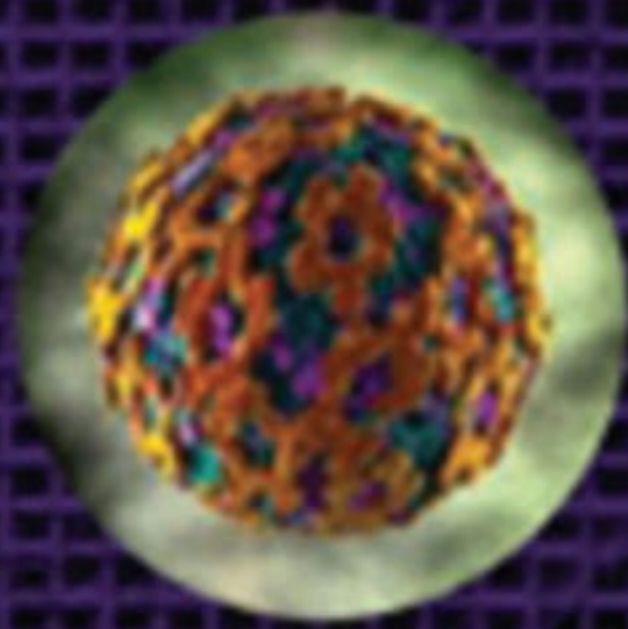


TENTH EDITION



HUNTER'S

Tropical Medicine *and*
Emerging Infectious
Diseases



EDWARD T. RYAN | DAVID R. HILL | TOM SOLOMON
NAOMI E. ARONSON | TIMOTHY P. ENDY

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Hunter's Tropical Medicine and Emerging Infectious Diseases

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Hunter's Tropical Medicine and Emerging Infectious Diseases

TENTH EDITION

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Edinburgh London New York Oxford Philadelphia St Louis Sydney 2020

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First edition 1945
Second edition 1954
Third edition 1960
Fourth edition 1966
Fifth edition 1976
Sixth edition 1984
Seventh edition 1991
Eighth edition 2000
Ninth edition 2013
Tenth edition 2020

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Chapter 38.5: West Nile Virus – by Marc Fischer, J. Erin Staples
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ISBN: 978-0-323-55512-8

Printed in Canada

Last digit is the print number: 9 8 7 6 5 4 3 2 1

Content Strategist: Charlotta Kryhl

Content Development Specialist: Nani Clansay

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Illustration Manager: Paula Catalano

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Preface

Hunter's Tropical Medicine grew out of the urgent need to provide training in tropical medicine to doctors preparing to support the massive American military expeditionary forces assembled during World War II. After America's official entry into the war in 1941, it was quickly apparent that millions of Allied military personnel would be deployed for years to tropical areas of the South Pacific, the China–Burma–India theater, North Africa, and, in smaller numbers, to South America and sub-Saharan Africa. The number of medical doctors who were knowledgeable and experienced in tropical infectious diseases to which they would be exposed was limited to a few specialized medical centers and the U.S. military. It became necessary to educate a large cohort of military medical corps officers very quickly. Much of this training was done at the Army Medical School tropical and military medicine course taught at the Army Medical Center in Washington, D.C.

The first edition of this book, entitled *Manual of Tropical Medicine*, was published in 1945 by three of the course instructors: Colonel Thomas T. Mackie, Major George W. Hunter III, and Captain Brooke Worth. The focus of the original book was a single small volume of practical information that could be carried by each doctor to his new assignment where they would be taking care of patients, often in relatively resource-poor environments. The same authors published a second edition in 1954. Colonel Hunter was joined by co-authors from Louisiana State University School of Medicine for the third, fourth, and fifth editions that were published in 1960, 1966, and 1976, respectively. George Hunter's contributions were acknowledged by adding his name to the book title in the sixth edition in 1984, edited by G. Thomas Strickland, a retired U.S. Navy captain. Dr. Strickland also edited the seventh and eighth editions published in 1991 and 2000.

Alan J. Magill, a retired U.S. Army colonel, took over for Dr. Strickland to lead the creation of the ninth edition published in 2013. Dr. Magill had a deep knowledge of tropical medicine, sharp clinical skills, leadership, and compassion. Dr. Magill passed away in 2015; at the time of his death he was director of the Bill & Melinda Gates Foundation malaria program. We dedicate this work to his lasting contributions in tropical medicine.

Although much has changed in the more than 80 years since the first edition was published, the current tenth edition of *Hunter's Tropical Medicine and Emerging Infectious Diseases* (HTM10) retains its primary objective as a concise presentation of practical information on the essential clinical aspects of patient presentation, diagnosis, and treatment of medical conditions found in the tropics. To accomplish this ambitious goal, a highly experienced and

dedicated group of editors has assembled a team of over 250 contributors from around the world. Numerous authors are from the tropics, and most of the authors who are not from the tropics have spent years living and working in the endemic areas.

We have produced a single volume of information with the clinician in mind, focusing on the perspective of a physician taking care of an individual ill patient. Tropical medicine has a long history dating back to the late 1800s, when the “germ theory” of disease was applied to the newly encountered diseases of the tropics as seen through the experience of European physicians sent out to new colonial destinations. For the purpose of HTM10, we use the term *tropical disease* as defined by the World Health Organization to specify a geographic area between the Tropic of Cancer (23.3 degrees latitude north) and Tropic of Capricorn (23.3 degrees latitude south). In practice, the term is often taken to refer to infectious diseases that thrive in hot, humid conditions, such as malaria, leishmaniasis, schistosomiasis, onchocerciasis, lymphatic filariasis, Chagas disease, African trypanosomiasis, and dengue, which are not endemic or are uncommon in temperate latitudes. Increasing human encroachment of tropical rainforests, deforestation, the impact of severe weather events and climate alterations, rising migration, international air travel, and tourism to and from tropical regions have led to an increased incidence and emergence of tropical diseases, including into temperate regions. Ebola, Zika, and chikungunya are just recent examples. Thus we added the title *Emerging Infectious Diseases* to the ninth and tenth editions of this series to underscore our global, interconnected, and constantly evolving reality.

It is our sincere wish that all who use this book find it useful in the care of individual patients and that the knowledge gained leads to improved outcomes for our patients and their communities.

Edward T. Ryan, MD

David R. Hill, MD

Tom Solomon, MD

Naomi E. Aronson, MD

Timothy P. Endy, MD

February 2019

Acknowledgments

The editors of the tenth edition of *Hunter's Tropical Medicine and Emerging Infectious Diseases* gratefully and sincerely acknowledge Dr. G. Thomas Strickland, the past editor of the sixth through eighth editions, and Dr. Alan J. Magill, the past editor of the ninth edition. Dr. Magill and Dr. Strickland worked tirelessly to create and update a clinically useful textbook of tropical medicine. We also extend a special thank you to all our contributors for the tenth edition, a highly talented and experienced group of clinicians assembled from around the world. We also sincerely thank the professional production and editing staff at Elsevier who worked so hard to complete this tenth edition. As individual editors, we would also like to honor, thank, and dedicate this work:

To my wife Krista; children Hana, Grace, and Edward; parents Edward and Ann; Thaire and Deborah; Dr. Sharon L. Ryan, family and teachers; my aunt Mary Theresa Dolan who died of an infection before her fifth birthday, and to all who have served, including Edward Ambrose Ryan, 22nd Marines, 1923–1944 (Kwajalein-Marshall Islands). The 22nd Marines were heavily affected by lymphatic filariasis following deployment in the Pacific Theater in 1942 and 1943. May the knowledge in this work be of use in advancing health and lessening suffering.

Edward T. Ryan, MD

To my parents and family who have constantly supported me, and to my students who will carry on the work of achieving global health equity.

David R. Hill, MD

To my six lovely girls: Rachel, Leah, Daisy, Rosie, Eva, and Peggy.

Tom Solomon, MD

To Jay and Lorraine Sanford for fostering inspiration.

Naomi E. Aronson, MD

To my family for their love and support.

Timothy P. Endy, MD

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SECTION A Organ-Based Chapters

1 Tropical Lung Diseases

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KEY FEATURES

- Pneumonia is a major cause of death in the tropics, especially in children under 5 years old.
- Symptoms and physical examination determine care-seeking behaviors and clinical management.
- Most new tuberculosis cases occur in the tropics, often with human immunodeficiency virus (HIV) co-infection.
- Parasitic infections can manifest as wheezing, eosinophilic pneumonia, pleural effusion, and cavitary lesions.
- The impact of non-communicable disease (e.g., chronic obstructive pulmonary disease [COPD] and lung cancer) on mortality is projected to rise in low- and middle-income countries.

INTRODUCTION

The term *tropics* refers to the region of earth lying between the Tropic of Cancer and the Tropic of Capricorn. In the tropics, warm climate, poverty, lack of education, and poor sanitation provide an ideal environment for pathogens, vectors, and intermediate hosts to flourish.¹ In this vast landmass, respiratory infections are a major cause of morbidity and mortality. Respiratory infections may be due to common pathogens encountered worldwide, as well as mycobacterial, parasitic, and fungal organisms. Infections are more prominent in immunocompromised hosts. On the other hand, non-communicable diseases (chronic obstructive pulmonary disease [COPD], chronic respiratory diseases, lung cancer, occupational lung diseases) are increasingly recognized in the tropics, where low socio-economic, educational, and nutritional status favor worse prognosis. A reasonable approach to the patient with lung disease in the tropics starts with age, occupational exposure, physical examination, HIV status, chest x-ray, and blood tests.

PNEUMONIA

World Health Organization (WHO) data showed that pneumonia is the leading cause of death in children under 5 years of age, and is responsible for 16% of deaths (Fig. 1.1). The incidence and mortality in the tropics are particularly increased, especially in South Asia and sub-Saharan Africa (Fig. 1.2). In Tanzania, 85% of children with suspected pneumonia are taken for care; however, proper care depends on family income, with the richest families seeking care 9.5 times more often. In Ethiopia only 30% of children with pneumonia were taken for care in a facility or in the community.² WHO data reveal that as few as 39% of newborns are breastfed, 60% of children with suspected pneumonia access care,

and 31% receive antibiotics. Poor vaccination, sanitation, and nutrition; crowded homes; indoor air pollution; and smoking all adversely affect pneumonia outcomes in the tropics.

Common pathogens include bacteria, viruses, and fungi. *Streptococcus pneumoniae* is the most common bacterial cause of pneumonia followed by *Haemophilus influenzae* type b. *Staphylococcus aureus* pneumonia accounts for 2% to 10% of acute community-acquired pneumonias.

Atypical pneumonias caused by *Mycoplasma pneumoniae*, *Chlamydia pneumoniae*, *Legionella* spp., viruses, fungi, and parasites occur. Respiratory syncytial virus (RSV) is the most common etiology of viral pneumonia. In HIV-infected infants, fungal pneumonia due to *Pneumocystis jirovecii* (formerly *P. carinii*) accounts for 25% of all pneumonia deaths. *M. pneumoniae* infections occur worldwide, affecting mostly school-aged children and young adults. The tropical physician should be aware of the non-respiratory manifestations of mycoplasma infection, including anemia, myringitis, Stevens–Johnson syndrome, hepatitis, and neuritis.

In 2002 to 2003, a coronavirus was responsible for more than 8000 cases of a severe acute respiratory syndrome (SARS) that spread via international travel across continents from its origin in Guangdong Province, China. After droplet inhalation of the virus, there was an incubation period of 2 to 7 days, then fever, cough, malaise, and headache. Pulmonary inflammation was characterized by desquamation of pneumocytes, hyaline membrane formation, and acute respiratory distress syndrome (ARDS). Recovery could be slow, and some patients developed fibrosis. Mortality was 10% to 20%, with the elderly and those with cardiovascular problems being especially at risk.

In 2012 another novel coronavirus appeared in Saudi Arabia, named *Middle East Respiratory Syndrome coronavirus* (MERS-CoV). Between 2012 and 2017, 2040 laboratory-confirmed cases have been reported to WHO, with small outbreaks outside of the Middle East. Transmission has occurred from human to human, but requires close contact and has particularly been seen in health care facilities between patients or between patients and health care providers. A dromedary camel reservoir is postulated. Initial symptoms are fever, cough, and shortness of breath, with disease severity ranging from asymptomatic or mild common cold symptoms to SARS. Thirty-five percent of confirmed cases have died.

Influenza viruses with pandemic potential are a risk for global disease spread.³ East Asia has seen continued outbreaks of avian influenza (e.g., H5N1, H7N9, and H9N2) with occasional spread to humans. In 2009–2010, a swine influenza virus (H1N1) originating in Mexico led to a global pandemic.

Investigations and Management

Sputum examination is an important aid in the diagnosis of pneumonia: color, amount, consistency, and odor. Mucopurulent sputum is commonly found in bacterial pneumonia or bronchitis. Scanty, watery sputum is often noted in atypical pneumonia; “rusty”

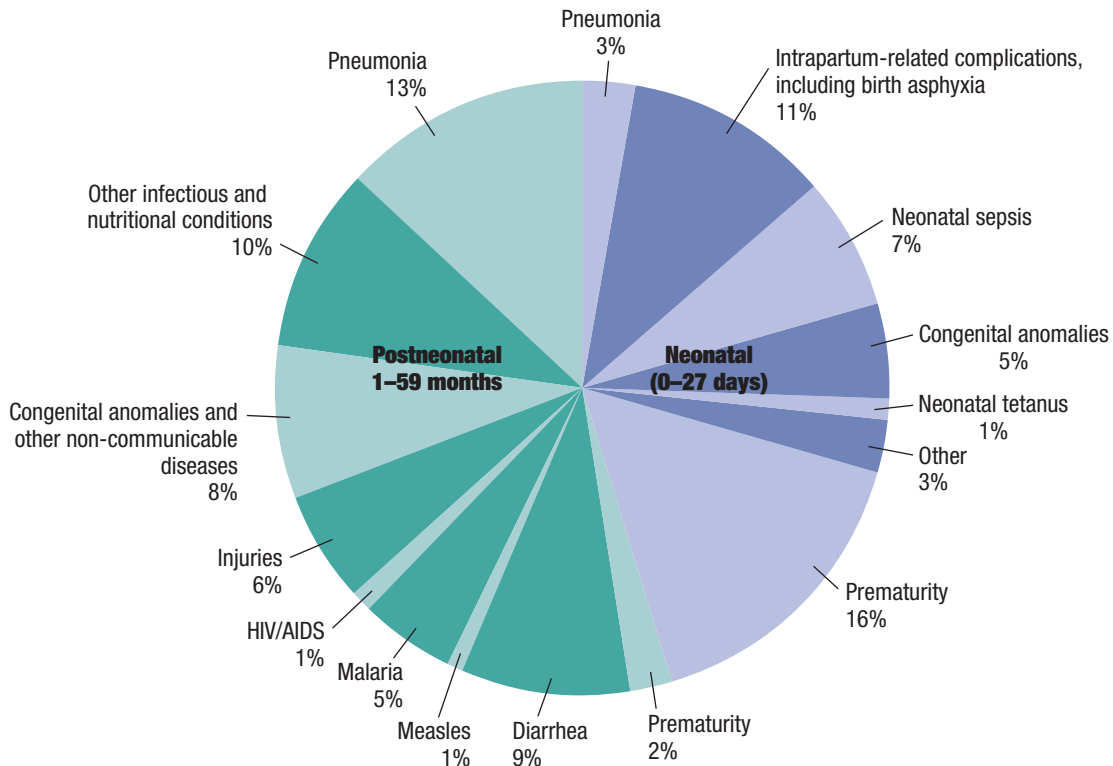


Fig. 1.1 Child causes of death, 2000-2015 (Global health estimates technical paper: WHO/HIS/IER/GHE/2016.1)

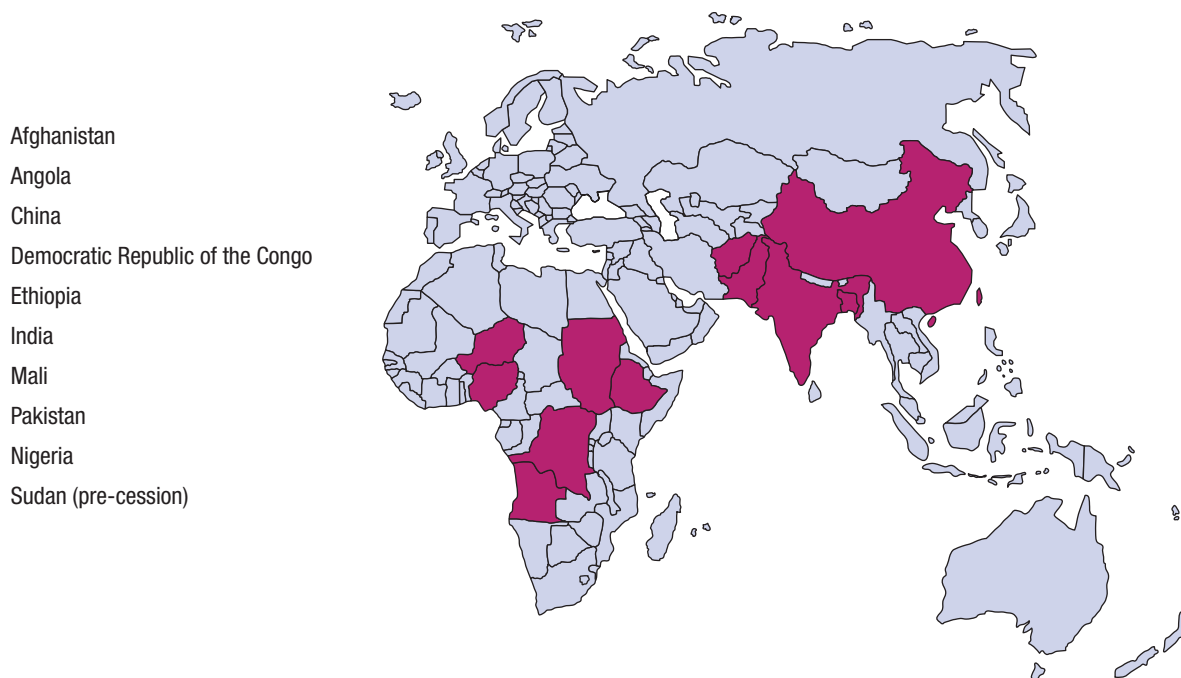


Fig. 1.2 Ten countries with the highest mortality from pneumonia. (Source: WHO Pneumonia and Diarrhea Report 2012; 2004 global burden of disease sub-analysis.)

sputum is seen in pneumococcal pneumonia; and currant-jelly or dark-red sputum suggests *Klebsiella pneumoniae*. Foul-smelling sputum is associated with anaerobic infections due to aspiration, lung abscess, and necrotizing pneumonia. A blood count usually reveals leukocytosis in bacterial pneumonia, a normal white cell

count or leukopenia in viral infection, and eosinophilia in parasitic infection. When available, chest x-ray, serum procalcitonin, and C-reactive protein can be obtained (Fig. 1.3). Naso-pharyngeal swabs for polymerase chain reaction (PCR) can help establish the diagnosis of specific viral infections if available.

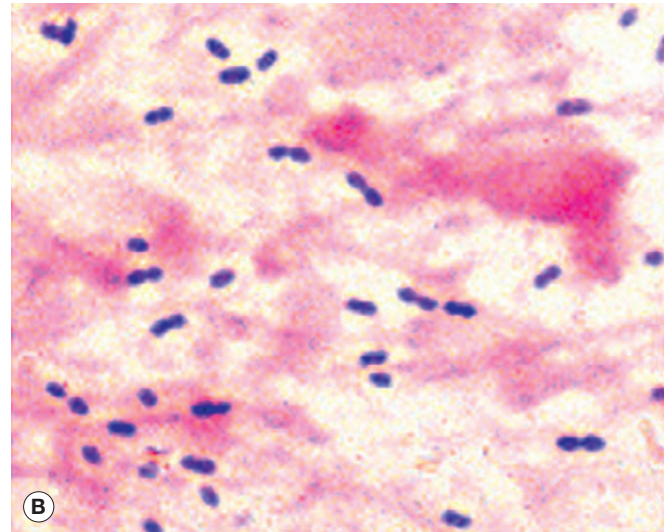
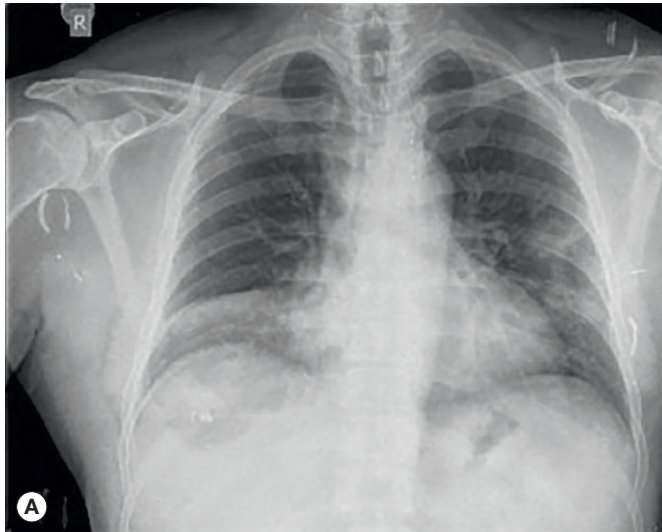


Fig. 1.3 (A) Pneumococcal pneumonia, right middle lobe. (B) Gram-positive diplococci (sputum).

TABLE 1.1 Revised WHO Classification and Treatment of Childhood Pneumonia at Health Facilities

1. Children with fast-breathing pneumonia with no chest in-drawing or general danger signs should be treated with oral amoxicillin: at least 40 mg/kg/dose twice daily (80 mg/kg/day) for 5 days. In areas with low HIV prevalence, give amoxicillin for 3 days. Children with fast-breathing pneumonia who fail on first-line treatment with amoxicillin should have the option of referral to a facility where there is appropriate second-line treatment.
2. Children age 2–59 months with chest in-drawing pneumonia should be treated with oral amoxicillin: at least 40 mg/kg/dose twice daily for 5 days.
3. Children aged 2–59 months with severe pneumonia should be treated with parenteral ampicillin (or penicillin) and gentamicin as a first-line treatment. Ampicillin: 50 mg/kg, or benzyl penicillin: 50,000 units per kg IM/IV every 6 hours for at least 5 days. Gentamicin: 7.5 mg/kg IM/IV once a day for at least 5 days. Ceftriaxone should be used as a second-line treatment in children with severe pneumonia having failed on the first-line treatment.
4. Ampicillin (or penicillin when ampicillin is not available) plus gentamicin or ceftriaxone are recommended as a first-line antibiotic regimen for HIV-infected and HIV-exposed infants and for children under 5 years of age with chest in-drawing pneumonia or severe pneumonia. For HIV-infected and HIV-exposed infants and for children with chest in-drawing pneumonia or severe pneumonia who do not respond to treatment with ampicillin or penicillin plus gentamicin, ceftriaxone alone is recommended for use as second-line treatment.
5. Empiric cotrimoxazole treatment for suspected *Pneumocystis jirovecii* (previously *P. carinii*) pneumonia is recommended as an additional treatment for HIV-infected and HIV-exposed infants aged from 2 months up to 1 year with chest in-drawing or severe pneumonia. Empirical cotrimoxazole treatment for *P. jirovecii* pneumonia is not recommended for HIV-infected and HIV-exposed children over 1 year of age with chest in-drawing or severe pneumonia.

In children, the Integrated Management of Childhood Illness guidelines for treating pneumonia are recommended (Table 1.1).⁴ Childhood pneumonia is now classified in two classes: tachypnea and chest in-drawing indicate treatment with oral antibiotics, whereas severe pneumonia with danger signs (hypothermia, unconsciousness, convulsions) requires intravenous treatment in hospital.

WHO's strategy to reduce pneumonia burden has three components: protection, by promoting breastfeeding and vitamin A supplementation; prevention, using vaccines for measles, *H. influenzae*, pertussis, rotavirus, and *S. pneumoniae* and promoting handwashing, sanitation, HIV prevention, and cotrimoxazole prophylaxis in HIV-infected patients; and treatment, securing care seeking, proper case management, and antibiotic administration.

TUBERCULOSIS

Tuberculosis (TB) is the most common infectious cause of death worldwide, surpassing both HIV and malaria.⁵ In 2015, 10.4 million incident cases were reported worldwide—480,000 with multi-drug resistant tuberculosis (MDR-TB).⁶ About half of these cases come from the tropics (India, Indonesia, Pakistan, South Africa, Nigeria) (Fig. 1.4). MDR-TB as well as extensively drug-resistant TB (XDR-TB) are increasing in African and Asian regions.⁷

Morbidity and mortality from TB increase with HIV co-infection and concurrence of diabetes mellitus. Refugee and migrant camps are hot-spots for TB transmission and spread. In Africa, the Ebola virus epidemic reduced TB as well as measles vaccination and favored disease spread. In Africa, 31% of TB cases have HIV co-infection, rising to 50% in regions of southern Africa.

Strategies to control TB are to monitor children under 5 years of age with exposure to TB cases, diagnose TB in HIV-infected subjects, implement modern diagnostic tools such as Xpert MTB/RIF that identifies the presence of TB and rifampin resistance, and make available anti-tuberculous drugs.

PARASITIC AND OTHER PULMONARY INFECTIONS IN THE TROPICS

These infections include malaria, pulmonary schistosomiasis, amebiasis, melioidosis, paragonimiasis, echinococcal cysts, Chagas disease, ascariasis, strongyloidiasis, filariasis, and tropical eosinophilia (Table 1.2).^{8–10} Individuals who come in contact with birds or animals may develop zoonoses such as tularemia, psittacosis, Q fever, and leptospirosis.

Leptospirosis is common in tropical areas where sanitation is poor and the water not adequately treated. Epidemics of leptospirosis occur after high rainfall in monsoon seasons when the water supply is contaminated by sewage or animal urine. About half of the patients with leptospirosis have fever, cough, hemoptysis,

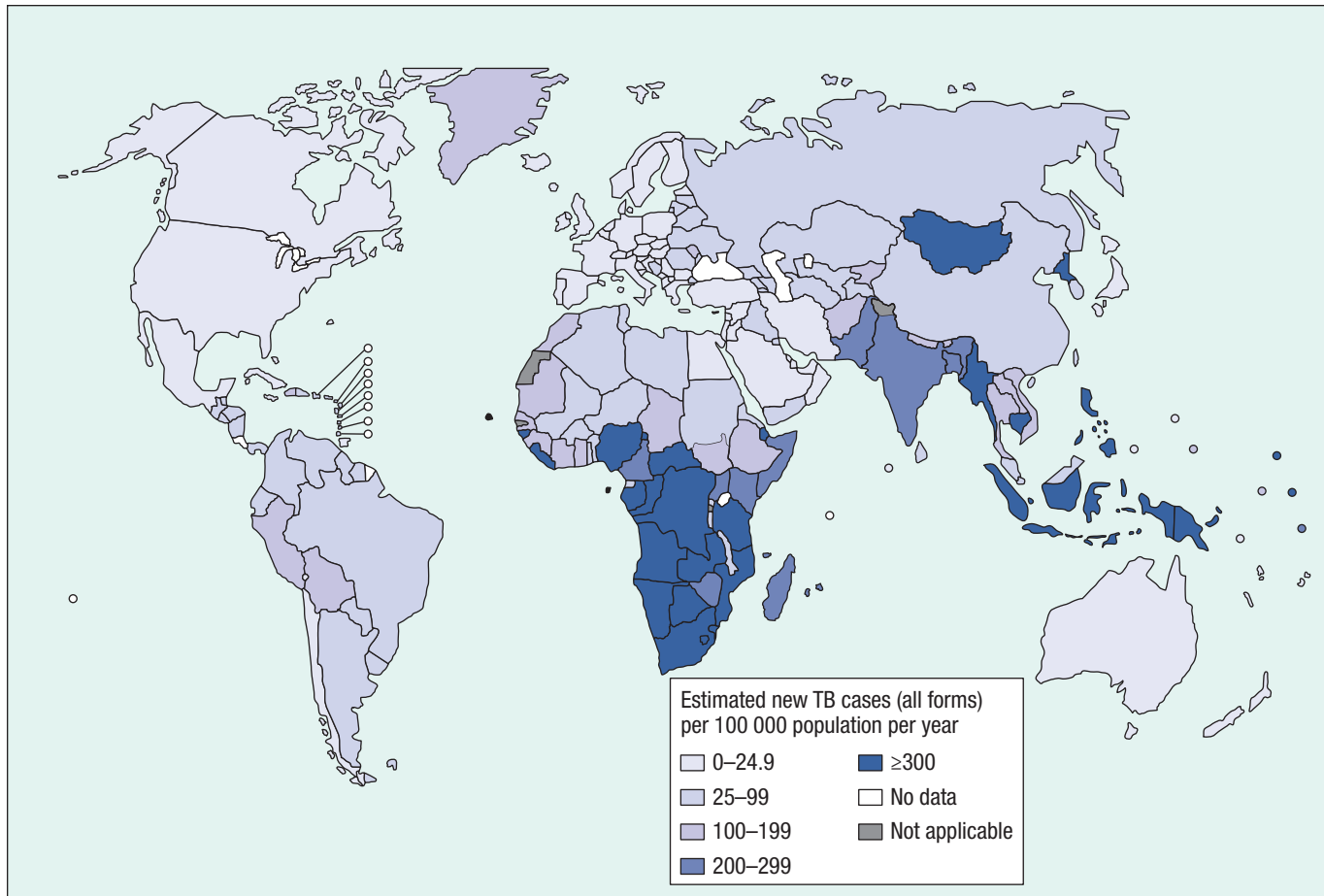


Fig. 1.4 Estimated TB incidence rates, 2015. (Source: WHO: 2016 Global Tuberculosis Report.)

TABLE 1.2 Key Pulmonary Manifestations of Parasitic Infections With or Without Eosinophilic Pneumonia

Schistosomiasis	Portal-pulmonary hypertension, arterio-venous fistulas
Amebiasis	Direct lung invasion through liver, pleuro-pulmonary involvement, anchovy sauce content
Malaria	Adult respiratory distress syndrome
<i>Trypanosoma cruzi</i>	Chagas disease
Ascariasis	Bronchopneumonia
<i>Strongyloides stercoralis</i>	Hyper-infection
Filariasis	Pulmonary infarction, milky pleural fluid
<i>Echinococcus</i>	Cyst, empyema, pneumothorax, hemoptysis, anaphylaxis
<i>Paragonimus westermani</i>	Eosinophilic pneumonia/pleural effusion
Melioidosis	Upper lobe cavity, acute/chronic infection, hemoptysis
<i>Wuchereria bancrofti</i> , <i>Brugia malayi</i>	Tropical pulmonary eosinophilia

and pneumonitis.¹¹ Other features are jaundice, conjunctivitis, and impaired renal function.

Melioidosis, caused by *Burkholderia pseudomallei*, is endemic in Southeast Asia (Vietnam, Cambodia, Myanmar), northern Australia, and West Africa. Melioidosis is hyperendemic in northern

Australia and in parts of northeastern Thailand and is an important cause of fatal community-acquired pneumonia. Chronic infection in patients with cystic fibrosis can occur.¹² The radiologic picture of upper lobe infiltration and cavity formation can be indistinguishable from TB. The mortality rate ranges from 20% to 50% but is higher in HIV-infected and immunocompromised hosts.

Respiratory symptoms of cough and chest pain in typhoid are present in up to 50% of cases at the onset of the disease. Pulmonary infiltrates may be associated with positive sputum cultures for *Salmonella typhi*. A fever chart showing continuous fever is highly suggestive of enteric fever. Diagnosis may be difficult without the ability to culture blood and stool.

In brucellosis, the lungs are involved in about 5% to 10% of cases, usually after inhalation of organisms. Abnormalities include bronchopneumonia, solitary or multiple lung nodes, miliary interstitial lung disease, lung abscess, and pleural effusion. Organisms can be identified on stains or sputum cultures.

Tularemia is a generalized infection caused by *Francisella tularensis* and occurs after skin or mucous membrane contact with infected mammals or through the bite of an arthropod, usually a tick or biting fly. Diagnosis should be considered when there is a skin ulcer associated with fever, generalized lymphadenopathy, cough, and signs of pneumonia. Pneumonia, either primary from inhalation of an infected aerosol or secondary to systemic infection, occurs in about 20% of cases.

Pneumonic plague is less common than either bubonic or septicemic disease. Nevertheless, fatal bronchopneumonia can occur without lymphadenopathy and is characterized by watery, bloody sputum. A sputum Gram stain can show bipolar stunted

rods. Pneumonic plague and tularemic pneumonia should be considered when a severe, rapidly progressive bronchopneumonia is reported in an endemic area and “typical” bacterial pneumonias have been ruled out.

In slaughterhouses, meat-processing plants, and areas with sheep and goat husbandry, Q fever (*Coxiella burnetii*) can cause epidemics of atypical pneumonia. Inhalation of dried infected material is the chief source; fever, headache, and dry cough are the main symptoms. Occasionally, the sputum is blood-streaked.

Bornholm’s disease (caused by coxsackie viruses and occasionally other enteroviruses), also known as *epidemic pleurodynia* or *devil’s grip*, causes chest pain and cough. Widespread epidemics of Bornholm’s disease occur in the Pacific islands and South Africa.

Kawasaki’s disease occurs in children under 5 years of age. This acute multi-system disease of unknown cause is characterized by fever of 5 days’ duration and four of five clinical features: non-purulent conjunctivitis; fissured lips, strawberry tongue; cervical lymphadenopathy; a maculopapular rash; and changes in the extremities with erythema and edema of the hands and feet. Pneumonitis occurs in 10% of the children and coronary artery dilatation and aneurysms in 20% to 25% of untreated cases. In Brazil, there has been a seasonal rise of the condition at the beginning and end of the monsoon season.¹³

Cryptococcus neoformans and *C. gatti* are saprophytic fungi distributed worldwide and are particularly abundant in soil in the tropics as well as in temperate countries. Pulmonary infection results from inhalation of the organisms from environmental sources. *C. neoformans* is the most common cause of meningoencephalitis in AIDS patients.

EOSINOPHILIC PNEUMONIAS

Systemic helminth infection usually elicits eosinophilia and increased levels of IgE. Although eosinophilia can be a clue to a pulmonary helminth infestation, the definitive diagnosis requires demonstration of ova or larvae in sputum, bronchial alveolar lavage fluid, pleural fluid, or lung biopsy.¹⁴ Loeffler’s syndrome refers to “simple” pulmonary eosinophilia with no or minimal systemic and pulmonary symptoms. In many helminth infections (*Ascaris*, strongyloidiasis, hookworm), the larvae migrate through the lung and in heavy infections can cause fever, cough, dyspnea, wheezing, hemoptysis, and lung infiltrate.

Schistosomes cause two clinical syndromes. In acute disease, immature schistosomula pass through the lung and can lead to fever, eosinophilia, and pulmonary infiltrate. In chronic schistosomiasis, especially when portal hypertension has led to venous shunts, eggs can bypass the liver and plug pulmonary capillaries and arterioles, producing granuloma and pulmonary hypertension. Radiographs may show dilated pulmonary arteries and arteriovenous abnormalities (Fig. 1.5).¹⁵

In paragonimiasis, the lung is the predominantly involved organ. The diagnosis must be considered in a patient from Southeast Asia with cough, hemoptysis (which is recurrent in >80% of cases), a pulmonary cavity, and pleural effusion.

Tropical pulmonary eosinophilia, typically in India and other South Asian countries, is caused by immunologic hyperresponsiveness to *Wuchereria bancrofti*, *Brugia malayi*, or other microfilariae. Clinical presentation consists of nocturnal cough, wheezing, fever, and weight loss. Chest radiographs show diffuse interstitial miliary infiltrates (Fig. 1.6); there is a high eosinophil count. In developed countries, serum IgE and anti-filarial antibodies can be used to confirm the diagnosis.

NON-COMMUNICABLE LUNG DISEASES

Non-communicable diseases (NCD) of the lung refer to non-infectious chronic respiratory diseases, such as COPD, bronchial asthma, lung cancer, and bronchiectasis.



Fig. 1.5 Bilateral pulmonary artery dilatation in schistosomiasis.

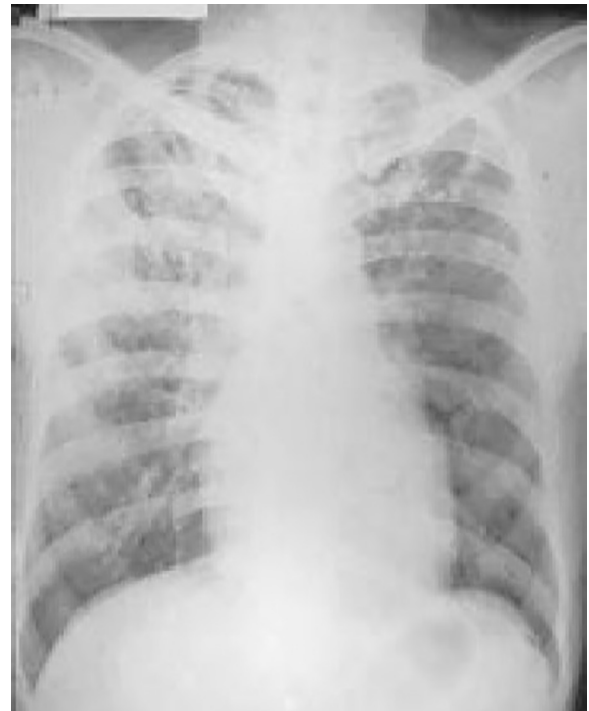


Fig. 1.6 Bilateral interstitial opacities affecting all lung fields in a patient with tropical pulmonary eosinophilia.

NCDs now represent an increasing portion of global morbidity and mortality burden in low- and middle-income countries. This is due to increased tobacco use and to low overall health and nutritional status, resulting in excess probability of mortality from NCD as shown in Fig. 1.7. In 2015 mortality in low-income countries accounted for 75% of overall global mortality. Most of the deaths occurred in South Asia and sub-Saharan Africa where mortality from NCDs and injuries often exceeded traditional infectious causes.^{16,17}

The incidence of asthma in the tropics is low for unclear reasons; however, “all that wheezes is not asthma” is a dictum that is true in the tropics, as there are many entities that cause wheezing and difficulty in breathing, including tropical eosinophilia and mitral

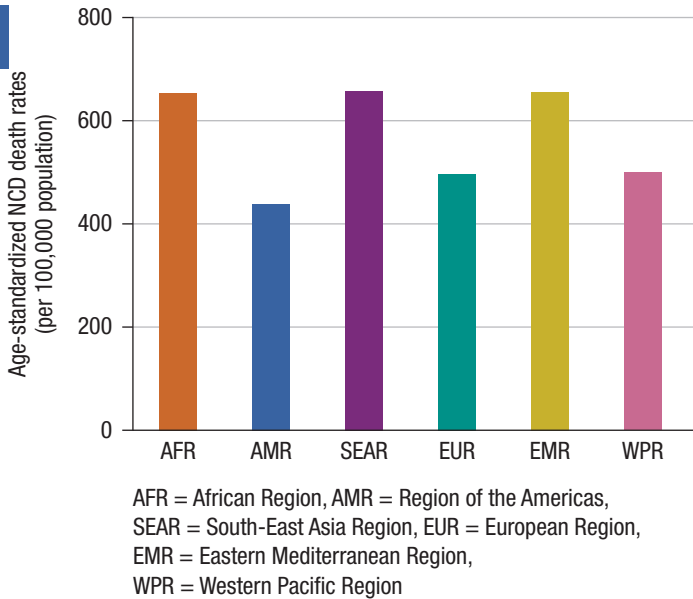


Fig. 1.7 High and increasing probability of mortality from non-communicable diseases in 2012. (Source: WHO: Global status report on non-communicable diseases 2014.)

stenosis. Asthma monitoring in the tropics can be achieved by using an inexpensive peak flow meter.

COPD is a progressive disease characterized by fixed airway obstruction. Causes of COPD in the tropics are increasing tobacco use and the widespread use of dung and biomass for indoor cooking and heating, demonstrated in one study in Indian males to decrease FEV₁ by 70 mL per year.¹⁸ The most common symptoms are dyspnea and chronic cough. The onset of dyspnea is insidious. With progression of airway obstruction, patients become short of breath at rest. When chest x-ray and pulmonary function testing are not available, a peak flow meter is an inexpensive device to assess severity of airway obstruction and monitor the response to treatment.

Bronchiectasis is a chronic, debilitating condition. Dilatation and distortion of the airways lead to impaired mucociliary clearance, which encourages bacterial colonization and bronchial inflammation. The diagnosis of bronchiectasis in developed countries is confirmed by computed tomography of the chest (Fig. 1.8), whereas in the tropics, the diagnosis is mainly clinical and depends on a compatible history, presence of finger clubbing, sputum that settles into three layers (mucoïd or frothy, mucopurulent, and purulent), and a chest x-ray, if available.

PLEURAL EFFUSION

Pleural effusion is frequent and has variable clinical signs and symptoms. Large effusions are associated with dyspnea and diminished chest movements on the affected side.

If possible, all but the smallest effusions should be tapped determining whether the fluid is serous, bloody, chylous, or contains pus. The effusion can be further divided into transudative and exudative, according to pleural fluid characterization. Laboratory tests that guide the management of a pleural effusion are macroscopic appearance, pleural fluid cell counts, biochemistry, pH, and Gram stain. A simple test is centrifugation of the fluid. If an originally “milky” fluid clears with that process, it is presumably an empyema. If not, it is either a chylothorax (pleural fluid triglycerides >110 mg/dL) or a cholesterol effusion (pleural fluid cholesterol >200 mg/dL).

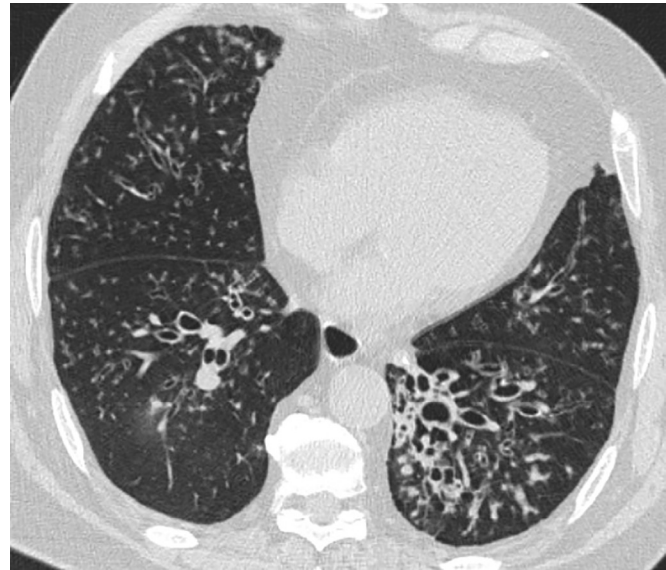


Fig. 1.8 Computed tomography of the chest: cylindrical and cystic bronchiectasis.

Transudative pleural effusions occur in heart failure, liver disease, endomyocardial fibrosis, hypoproteinemia/malnutrition, and hypothyroidism. The pleural fluid white blood cell count is typically <1000 cells/mm³, the pH >7.2, protein <3.0 g/L, the lactate dehydrogenase (LDH) <200 IU/L, and the glucose ≥60 mg/dL. A bloody effusion is caused by hemothorax, trauma, malignancy, and pulmonary embolism.

Exudative effusions typically have cell counts, protein, and biochemical markers opposite to those of transudates. Exudates can be further classified into neutrophilic, lymphocytic, and eosinophilic. A neutrophilic exudate occurs commonly in bacterial pneumonia cases and can progress to a complicated effusion or to an empyema, both necessitating pleural fluid drainage with a chest tube thoracostomy in addition to antibiotic treatment.

The disease presenting with the highest pleural fluid lymphocytosis is tuberculous pleuritis. A large volume of pleural fluid should be obtained for examination for acid-fast bacilli. In about one-third of cases, the tuberculin skin test is negative initially and converts to positive after 2 to 4 weeks.

An eosinophilic exudate is more common in the tropics. Endemic parasitic and fungal infections are major causes of such an effusion. Ascariasis, echinococcosis, and paragonimiasis are some of the parasitic infections. Paragonimiasis is associated with low pleural fluid glucose and low pH. Fungal diseases responsible for such an effusion are histoplasmosis, cryptococcosis, and coccidioidomycosis.

NON-TUBERCULOUS GRANULOMATOUS LUNG DISEASE

In the absence of chest x-ray or biopsy evidence, it is not possible to diagnose pulmonary involvement due to sarcoidosis and other granulomatous diseases. Consequently, in the tropics, these disorders usually remain undiagnosed. The possibility of sarcoidosis should be considered in a patient with dyspnea, uveitis, hepatosplenomegaly, peripheral lymphadenopathy, chronic skin lesions, and a chest x-ray film showing bilateral hilar adenopathy.¹⁴

OCCUPATIONAL AND DUST LUNG DISEASES

The occupational disorders result from human social activity, and as such are preventable. The dusts that provoke occupational

disorders can be classified into those that induce granulomatous reaction (e.g., beryllium, talc and organic antigens); those that cause fibrosis (e.g., silica, asbestos, and coal); and those that cause neither inflammation nor fibrosis, thus remaining inert (e.g., iron, barium, and tin). Poorly recognized occupational diseases in the tropics are byssinosis (due to cotton dust), mostly in Asia and Africa; bagassosis (due to sugar cane), mostly in Americas, Cuba, and India; and hypersensitivity pneumonitis.

Podoconiosis is an endemic non-filarial elephantiasis occurring in individuals exposed to red clay soil derived from alkaline rock. The silica particles are found in the skin, lymph nodes, and lymphatics of affected and unaffected individuals. These individuals have reduced lung function compared with adults living in areas of low silica concentration.¹⁹

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